Vector-Borne Blood Protozoa in Cattle and Sheep of Identification Mahmudul Hasan¹, Mamunur Rahman², Ali Khan^{3*}

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Abstract

Babesiosis, animal disease and Theileriais square measure the foremost common vector (Tick) borne blood protozoan diseases (TBDs) in People's Republic of Bangladesh. This study was conducted in kine and sheep during a completely different space of People's Republic of Bangladesh. a complete range of 1150 blood samples were indiscriminately collected from capital of Bangladesh, Sirajganj and Nikhangsori for blood smear research. However, co-infections, temperature, humidity, season, farming and prevention were additionally into consideration. From the clinically positive sample PCR was done followed by gel ionophoresis. Prevalence of blood protozoa were 100 percent (55), eightieth (n=320), half-hour (n=120), 22% (n=44), 31% (n=22), sixty fifth (n=16) in exotic sheep, intensive farming, milk-vita space, native kine, hill tracts and native sheep severally. the general prevalence was fifty.17% (n=577). Among the protozoa, Anaplasma spp. was 43%, Babesia spp. 19%, Anaplasma spp. with Babesia spp. 33%, Theileriaspp fourdimensional and Anaplasma spp. with Babesia and Theileriaspp was 1 Chronicles. The prevalence of blood protozoa in native breed ≥50%, up to seventy fifth and higher than seventy fifth cross or pure breed were seventeen.58% (n= 103), 31.91% (n=187) and fifty.51% (n=296) severally. Prevalence of blood protozoa throughout Oct to March was sixteen.041% (n= 94) and Gregorian calendar month to Sep was eightythree.959% (n=492). In PCR Anaplasma marginale showed positive band as 265 bp, Babesiabovis in 166 bp, and Theileriaannulata in 312 bp, Babesiaovis in 422bp and Babesiamotasi in 518bp severally. Therefore, the tick is act as vector and high wetness and temperature is that the main risk issue for vector borne diseases. finally, blood protozoa square measure the silent rising unwellness in eutherian mammal and want to boost the management strategy.

Key words: Anaplasmaspp; PCR; Prevalence; Vector.

INTRUDUCTION

Anaplasma spp may be a gram-negative bacterium protozoan whereas Babesiaspp, and Theileriaspp area unit apicomplexan parasite that infects red corpuscle (RBC), so transmission happens to animal through vector bite, notably Ixodes and frequently grasp as tick borne diseases (TBDs) (Karim et. al., 2012) and conjointly worldwide distributed [1,2]. interdependency one among the main hinders in placental mammal farming in Bangladesh and hot wet climatical condition greatly favours the event and survival of ecto and endo parasite that creates the violence of interdependency and is aware of as disease [3]. Crossbreed animals were a lot of

vulnerable than autochthonal cows and summer season was predominant for blood protozoa followed by winter and season in tropical and sub-tropical countries. Adult and feminine were a lot of vulnerable than young and male [4]. The clinical sign showed that prime fever (105-1070F), anaemia, riotous diarrhoea, ascites, someday bloody diarrhoea, low color pee ultimately stage of inBabesiosis [5]. regarding eightieth of the planet cow's population is suffering from TBDs [6]. The TBDS in Bangladesh predominated in forest and craggy areas, high humidness and temperature irritate the outbreaks and cross or pure animal breed is a lot of prone to infection [7]. wet and hot climatical condition favors the expansion, multiplication and survival of tick and blood protozoa in

erythrocyte that causes a deadly disease of TBDs [8]. It causes anaemia, hides injury, reduces milk production and poor fruitful performance, enhanced mortality and international economic losses calculable at US\$ eighteen. 7 billion [9]. In blood smear research, Babesia spp. tally as short and long loop formation in erythrocyte (Piroplasmosis), Anaplasmamarginale like as pointed spherical dot at boundary of erythrocyte and in Anaplasmacentrale pointed spherical dot inside erythrocyte. wherever as in Theileriaspp erythrocyte was ringed and, round, dot, rod form was found [10]. Biting flies transmit the malady and multiplication is enhanced in sexual stages thanks to hot wet environment [11]. typically recovered animal act as persistent carriers. ancient impression smear staining may be a routine check want to determine Babesia, Theileria and Anaplasma spp. enzyme chain reaction (PCR) is currently turning into a typical tool for molecular detection [12]. Recovered animal act as carrier and notably, making a possible supply of infection [13]. The multiplex PCR with species specific primer gave positive bands at 166bp, 265bp and 312bp selective for B. bovis, A. marginaleand T. annulata in cows severally [14]. However, Anaplasma marginale, Anaplasmacentral, BabesiaOvis, Babesiamotasiand Theileriaannulataare the cause for TBDs in sheep [15]. Early diagnosing and specific treatment in conjunction with vector management area unit necessary to forestall death and production losses. Therefore, considering the importance of TBDs the analysis work was in deep trouble the identification and molecular detection of vector borne blood protozoan infection notablyBabesiaspp, Anaplasmaspp and Theileriaspp in cows and sheep in Bangladesh with seasonal variation.

MATERIALS AND METHODS

Study Area

A medicine study was administrated in Parasitology Laboratory below Associate in Nursingimal Health analysis Division in People's Republic of Bangladesh stock analysis Institute (BLRI), Savar, capital of Bangladesh from July 2016 to June 2017.

Sample Assortment

About 2.5 cc of peripheral blood samples were collected from completely different kine and sheep farm from many of Savar, SirajganjSadar, ShajadpurUpazila and Nikhangsori, Chottrogram at intervals gas organic compound characin fish Acetate (EDTA) tube with icecool storage and shifted to the Parasitology laboratory in BLRI. a complete variety of 1150 blood samples were collected indiscriminately on form basis, among them fifty five from Australian sheep (BLRI), four hundred from a high yielding farm of Savar, four hundred from high yielding kine from milk vita Bathan space (Baghabari), two hundred from native kine (Sirajganj), seventy from native mountainous kine of Nikhangsori, twenty five from native sheep.

Laboratory Identification of Blood Protozoa through Giemsa's Stain Methodology

Samples were examined by Giemsa's stained blood smear (GMS) research (FAO, 2016) and substantiating designation through enzyme Chain reaction (PCR). The result of topography, season, age and sex was remaining in thought during this study. In GMS protocol thick and skinny blood smear was done. when air dry absolute methyl alcohol fixation was done and stained with 100 percent Giemsa's stain. when laundry, air dry and emulsification, magnification below 100x objectives. The haemoprotozoa were microscopically known supported the characteristic morphology illustrated by Soulsby.

DNA Extraction of Blood Protozoa

Blood protozoan polymer was extracted employing a commercially on the market kit (Invitrogen Purelink Genomic polymer mini kit, Cat. no. K1820-01) from blood sample through chloroform methodology. At a look, two hundred two hundred blood was mixed with two hundred two hundred of lysis buffer containing 20µg/ml protease K and incubation was done at fiftyfive 0C for ten minutes. Thereafter, laundry and natural process was done at thirteen,000x g for three minutes. Finally, the spin column was discarded and aggregation the eppendorf tube containing extracted polymer and keep in -20 0C within the icebox. The purity of genomic was envisioned by victimisation spectrophotometry (260°A/280°A) with one.5% gel dielectrolysis (Sigma Aldrich, USA). The compaction of the polymer ordering was adjusted to $100 \text{ng}/\mu\text{l}$ enzyme free water.

Multiplex Enzyme Chain Reaction (PCR)

PCR was administrated during a final reaction volume of 25µl within the skinny walled PCR tubes to amplify genomic polymer of Babesia, Anaplasma and Theileria species. The commercially on the market master combine kit (Thermo Scientific) was wont to amplify fragments of genomic polymer during a programmable thermocycler (Eppendorf, Germany). what is more, when Associate in Nursing initial catalyst activation step at 95°C for five min, the reaction mixture was subjected to thirty five cycles every containing a denaturation step at 95°C for thirty sec, Associate in Nursing hardening step at 68°C for thirty sec, Associate in Nursingd an extension step at 72°C for 1.5 min. when a final elongation step at 72°C for five min, PCR product were resolved by agarose gel dielectrolysis, stained with ethidium bromide, so ascertained below ultraviolet|ultraviolet illumination|UV|actinic radiation|actinic ray} light.

Oligonucleotide primers were utilized in the PCR amplification cycle (First BASE Laboratories sdnbhd, Malaysia). The PCR pictures were captured although pc software system (Carl Zeiss, GmbH, Germany) and therefore the positive samples were detected by specific band size of the PCR product.

RESULTS AND DISCUSSION

In blood smear microscopy prevalence of TBDs was 100% (n=55) in Australian sheep, 80% (n=320) in dairy farm, 30% (n=120) in Bathan area, 22% n= (44) in native cattle, 31% (n=22) in hilly cattle, and 65% (n=16) in native sheep, this findings strongly supported where

he had found that prevalence of TBDs was significantly varied on area, season and breed.

The overall prevalence of TBDs was 50.17% (n=577) in cattle and sheep in which Anaplasma spp was 43%, Babesia spp 19%, Anaplasma spp. and Babesiaspp 33%, Theileriaspp 4% and Anaplasma spp. with Babesia and Theileriaspp 1% of blood protozoa (Table 3). This result was almost similar with where stated that in Turkey, the overall prevalence was 74.78%, Anaplasma spp and Babesia spp was 41.99%, and slightly higher from where over all prevalence was 38%, there was some variation due tropical and subtropical regions variation. In positive case the blood protozoa magnify slight purple color. In case of Babesia spp. short and long loop formation was found at the periphery of RBC (Piroplasmosis). In case of Anaplasma marginale pointed round dot at periphery of RBC and in Anaplasmacentrale pointed round dot inside of RBC. In case of Theileriaspp RBC was slight triangle in shape and ring form Theileriaspp was found (annular), sometimes oval, round, dot, rod shape was found, this finding was notably similar with (Figure 1).

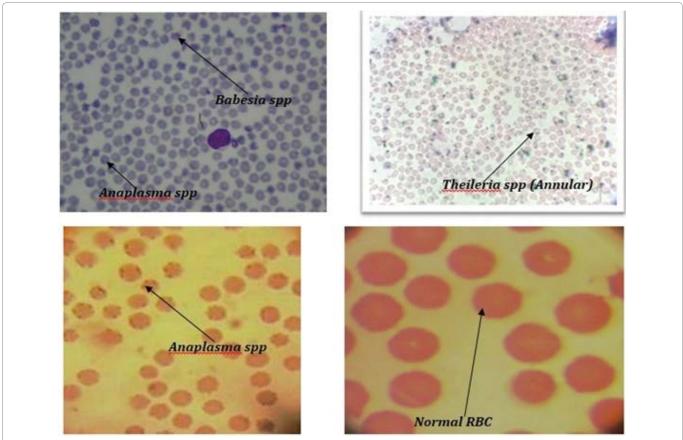
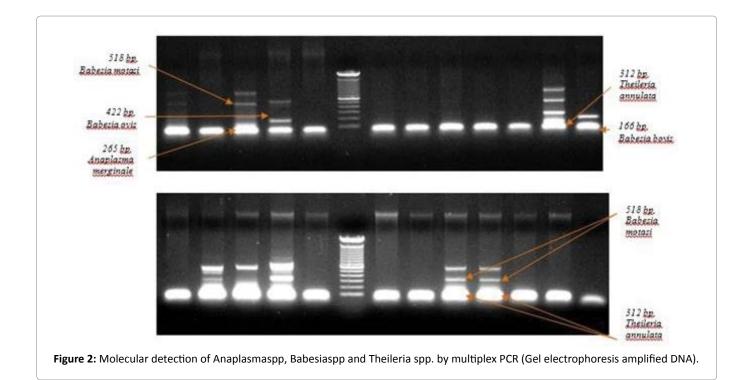


Figure 1: Blood smear microscopy of Babesiaspp, Anaplasmaspp and Theileriaspp within RBC in 100x objectives.



However, Anaplasma marginaleshown positive band as 265 bp, Babesiabovisin 166 bp, and Theileriaannulata in 312 bp, in cattle blood whereas Anaplasma marginalein 265 bpBabesiaovisin 422bp, Babesiamotasiin 518bp and Theileriaannulata in 312 bp in sheep blood respectively, this finding was clearly significant with (Figure 2).

However in seasonal study in Bangladesh it was observed that April to September environmental temperature is arise (above 300°C, sometimes 400°C) and humidity is above 70% (sometimes above 90%) that triggers the multiplication of tick biologically and also multiplication of TBDs protozoa both in tick and animal blood that progresses havoc of TBDs in high yielding animal and local animal act as carrier, this clarification also clearly justified the same findings, there he stated that infection in sheep 52% due to hot humid environmental condition.

In addition, when environmental temperature is 300C or below and humidity is below 70% notably during October to March animal act as carrier but not showing clinical sign, and prevalence of blood protozoa during October to march was 16.041% (n= 94) and April to September was 492 83.959% (n=492), this result was verified the findings.

In case of high yielding animal (above 60% cross breed) and 100% pure breed show high clinical sign and even death in high percentage and response to treatment is

low. In local breed or 50% crossed breed, upto 75% crossed breed and above 75% or even pure prevalence of blood protozoa was 17.58% (n= 103), 31.91% (n=187) and 50.51% (n=296) respectively. The findings compared with where signifying that the crossbred is more susceptible to TBDs than local animal and consequently, strongly supported by where they were stated that, TBDs caused high morbidity, mortality and economic losses in high yielding animal than local breed of ruminants.

CONCLUSION

Tick borne blood protozoan disease (Babesiosis,AnaplasmosisandTheileriosis) are now a days a crucial factor for livestock production in Bangladesh. Local animal act a as carrier but it is indicating future havoc in livestock industry especially high yielding exotic animal (70 % to 100 % pure breed). Moreover, they are more susceptible to TBDs and it is very difficult to control because high temperature and humidity provoke the tick multiplication. To introduce high yielding animal in a farm strict biosecurity is essential for farming.

REFERENCES

1. Vieira LL, Canever MF, Cardozo LL, Cardoso CP, Herkenhoff ME, Neto AT, et al. Prevalence of Anaplasma marginale, Babesia bovis, and Babesia bigemina in cattle in the Campos de Lages region, Santa Catarina state, Brazil, estimated by multiplex-PCR. Parasite Epidemiol Control. 2019 Aug; 6: e00114.

- 2. https://www.semanticscholar.org/paper/Development-ofmultiplex-PCR-assay-for-concurrent-Kundave-Ram/137e8178c2a ef824ef110b7717648c03c507e1c2
- 3. https://curis.ku.dk/ws/files/49318138/Roy_Prevalence_of_ hemoprotozoan_diseases_in_cattle_population_of_chittagong_ division_Bangladesh.pdf
- Aouadi A, Leulmi H, Boucheikhchoukh M, Benakhla A, Raoult D, Parola P. Molecular evidence of tick-borne hemoprotozoanparasites (Theileria ovis and Babesia ovis) and bacteria in ticks and blood from small ruminants in Northern Algeria. Comp Immunol Microbiol Infect Dis. 2017 Feb;50:34-39.
- 5. http://pvj.com.pk/pdf-files/32_2/221-224.pdf
- 6. Ghosh S, Azhahianambi P, Yadav MP. Upcoming and future strategies of tick control: a review. J Vector Borne Dis. 2007 Jun;44(2):79-89.
- Bhat S, Singh N, Singh H, Rath S. Molecular prevalence of Babesia bigemina in Rhipicephalus microplus ticks infesting cross-bred cattle of Punjab, India. Parasite Epidemiol Control. 2017 Aug; 2(3): 85–90.
- 8. Lee SH, Mossaad E, Ibrahim AM, Ismail AA, Adjou Moumouni PF, Liu M, et al. Detection and molecular characterization of tick-borne pathogens infecting sheep and goats in Blue Nile and West Kordofan states in Sudan. Ticks Tick Borne Dis. 2018 Mar;9(3):598-604.

- 9. Krishna murthy CM, Ananda KJ, Adeppa J. Prevalence of Haemoprotozoan infections in bovines of Shimoga region of Karnataka state. J Parasit Dis. 2016 Sep; 40(3): 890–892.
- 10. https://onlinelibrary.wiley.com/doi/full/10.1111/tbed.12894
- 11. Ghosh S, Nagar G. Problem of ticks and tick-borne diseases in India with special emphasis on progress in tick control research: a review. J Vector Borne Dis. 2014 Dec;51(4):259-70.
- 12. Bock R, Jackson L, de Vos A, Jorgensen W, Babesiosis of cattle. Parasitology. 2004;129 Suppl:S247-69.
- 13. Zhou M, Cao S, Sevinc F, Sevinc M, Ceylan O, Ekici S, et al. Molecular detection and genetic characterization of Babesia, Theileria and Anaplasma amongst apparently healthy sheep and goats in the central region of Turkey. Ticks Tick Borne Dis. 2017 Feb;8(2):246-252.
- Kocan KM, de la Fuente J, Blouin EF, Garcia-Garcia JC. Anaplasma marginale (Rickettsiales: Anaplasmataceae): recent advances in defining host-pathogen adaptations of a tick-borne rickettsia. Parasitology. 2004;129 Suppl:S285-300.
- 15. Bilgiç HB, Karagenç T. Development of a multiplex PCR assay for simultaneous detection of Theileria annulata, Babesia bovis and Anaplasma marginale in cattle. Exp Parasitol. 2013 Feb;133(2):222-9.